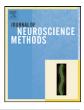


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The use of motion analysis to measure pain-related behaviour in a rat model of degenerative tendon injuries

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ABSTRACT

Chronic tendinopathy is characterized with longstanding activity-related pain with degenerative tendon injuries. An objective tool to measure painful responses in animal models is essential for the development of effective treatment for tendinopathy. Gait analysis has been developed to monitor the inflammatory pain in small animals. We reported the use of motion analysis to monitor gait changes in a rat model of degenerative tendon injury. Intratendinous injection of collagenase into the left patellar tendon of Sprague Dawley rat was used to induce degenerative tendon injury, while an equal volume of saline was injected in the control groups. Motion analyses with a high speed video camera were performed on all rats at pre-injury, 2, 4, 8, 12 or 16 weeks post injection. In the end-point study, the rats were sacrificed to obtain tendon samples for histological examination after motion analyses. In the follow-up study, repeated motion analyses were performed on another group of collagenase-treated and saline-treated rats. The results showed that rats with injured patellar tendon exhibited altered walking gait as compared to the controls. The change in double stance duration in the collagenase-treated rats was reversible by administration of buprenorphrine (p = 0.029), it suggested that the detected gait changes were associated with pain. Comparisons of end-point and follow-up studies revealed the confounding effects of training, which led to higher gait velocities and probably a different adaptive response to tendon pain in the trained rats. The results showed that motion analysis could be used to measure activity-related chronic tendon pain.

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1. Introduction

Chronic tendinopathy refers to insidious onset of chronic activity-related pain that can affect virtually all tendons. The pathogenesis of chronic tendinopathy remains speculative (Riley, 2004). It is generally regarded as a result of failed healing to accumulated micro-injuries (Khan et al., 1999). Treatments are empirical and symptom-based, and the responses to treatment vary a lot (Alfredson, 2005). Previous experimental studies on chronic tendinopathy included analyses of clinical samples of tendinopathic tissues (Fu et al., 2002a,b, 2007; Rolf et al., 2001). Histopathological changes similar to those in clinical samples of tendinopathy were reproduced in animal models by overuse (Glazebrook et al., 2008), injection of collagenase (Chen et al., 2004) or cytokines (Stone et al., 1999). However, owing to a lack of measurement of pain, representative animal models for chronic tendinopathy could not be

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established. Breakthroughs for experimental studies for chronic tendinopathy will reside on the development of an objective measure of tendon pain in animal models.

A number of methods were developed to measure pain in small animals, including measurement of weight-bearing (Vrinten and Hamers, 2003), mechanical sensitivity (Fernihough et al., 2004), vocalization (Han et al., 2005) and gait analysis such as footprint analysis (Marxen et al., 2004) and motion analysis (Coulthard et al., 2002, 2003; Varejao et al., 2002). As chronic tendinopathy is characterized with activity-related pain, it is more plausible to detect the pain-associated changes in dynamic state (walking) instead of static state (standing). Gait analysis has been developed to monitor inflammatory pain in small animals such as rats (Coulthard et al., 2003), as well as large animals such as horse (Marxen et al., 2004). Coulthard et al. (2002, 2003) demonstrated the use of motion analysis as a reproducible, objective measure for acute and chronic pain induced with intraplantar injection of irritants, characterized by significant changes in temporal gait parameters such as double stance duration. Changes in ankle angle during stance phase have been used to investigate functional recovery in rats (Varejao et al., 2002). Knee joint pain induced by arthritis was widely studied in animal model by motion analysis (Neugebauer et al., 2007), but

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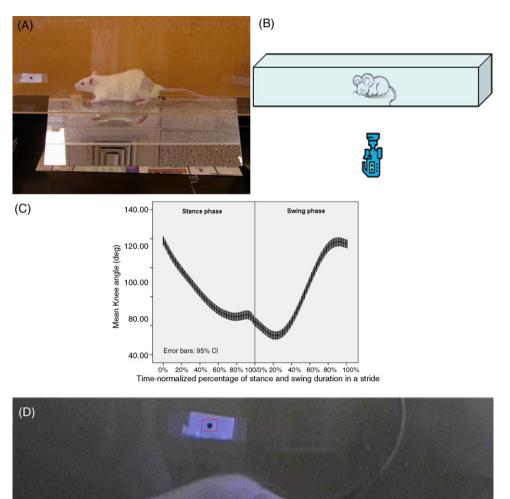


Fig. 1. Experimental set-up of motion analysis on the walking gait of rats. The rats were encouraged to walk through a transparent walkway (A) during motion analysis. The video camera was positioned to capture the rat motion in the saggital plane as shown in the diagram (B). Both hind limbs of the rats were marked on the joints of hip, knee, ankle and toe and checked for consistency of marks and joint movement by moving the limbs, as shown by a consistent kinematic plot for knee angle during a stride (C). With the help of the reference point and the marks on the joints, the gait parameters and knee angles were measured by a motion analysis system (D).

it is still unexplored if pain associated with degenerative patellar tendon injury could be monitored by the same technique.

In the current study, we reported the use of motion analysis of the saggital plane of walking gait to measure painful responses in a rat model of collagenase-induced degenerative tendon injury. Several unexplored areas in measurement of pain-associated gait changes were addressed. Firstly, changes in the contralateral side of the injured limb were analyzed in order to evaluate if there was compensatory change in the contralateral side. Secondly, two different double stance durations within a stride were separately measured. Double stance duration (DS) is defined the duration when both limbs touch the ground during a stride. It follows that two separate DS are identified in a stride: one ended with the take-off of the contralateral limb, while another ended with the take-off of the observed limb. However, there was no clear distinction between the two in previous reports (Coulthard et al., 2002, 2003). Since DS is identified as a sensitive parameter to characterize pain (Simjee et al., 2004), it is necessary to distinguish the two for the use of DS as a valid measurement for pain. Thirdly, the

influences of repeated measurements as training effect were evaluated when comparing the results of follow-up study with those of end-point measurement.

2. Materials and methods

The procedures in the following animal experiments were approved by the Animal Research Ethics Committee, the Chinese University of Hong Kong.

2.1. Experimental design

Fifty-eight male Sprague Dawley rats, at an age of 8 weeks, weighting 250-300 g, were used in this study. In the end-point study, forty-two rats were randomly assigned into different groups. Six rats were subject to motion analysis without injection to the knee as pre-injection group. Thirty rats were assigned to receive collagenase injection and subject to motion analysis at 2, 4, 8, 12 and 16 weeks post injection (n=6). Six rats received saline

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.jneumeth.2009.02.011.

References

- Alfredson H. Conservative management of Achilles tendinopathy: new ideas. Foot Ankle Clin 2005;10(2):321–9.
- Chen YJ, Wang CJ, Yang KD, Kuo YR, Huang HC, Huang YT, et al. Extracorporeal shock waves promote healing of collagenase-induced Achilles tendinitis and increase TGF-beta1 and IGF-I expression. J Orthop Res 2004;22(4):854–61.
- Coulthard P, Pleuvry BJ, Brewster M, Wilson KL, Macfarlane TV. Gait analysis as an objective measure in a chronic pain model. J Neurosci Meth 2002;116:197–213.
- Coulthard P, Simjee SU, Pleuvry BJ. Gait analysis as a correlate of pain induced by carrageenan intraplantar injection. J Neurosci Meth 2003;128:95–102.
- Fernihough J, Gentry C, Malcangio M, Fox A, Rediske J, Pellas T, et al. Pain related behaviour in two models of osteoarthritis in the rat knee. Pain 2004;112:83–93.
- Filipe VM, Pereira JE, Costa LM, Mauricio AC, Couto PA, Melo-Pinto P, et al. Effect of skin movement on the analysis of hindlimb kinematics during treadmill locomotion in rats. J Neurosci Meth 2006;153(1):55–61.
- Fu SC, Chan BP, Wang W, Pau HM, Chan KM, Rolf CG. Increased expression of matrix metalloproteinase 1 (MMP1) in 11 patients with patellar tendinosis. Acta Orthop Scand 2002a;73(6):658–62.
- Fu SC, Chan KM, Rolf CG. Increased deposition of sulfated glycosaminoglycans in human patellar tendinopathy. Clin J Sport Med 2007;17(2):129–34.
- Fu SC, Wang W, Pau HM, Wong YP, Chan KM, Rolf CG. Increased expression of transforming growth factor-beta1 in patellar tendinosis. Clin Orthop Rel Res 2002b;400:174–83.
- Glazebrook MA, Wright JR, Langman M, Stanish WD, Lee JM. Histological analysis of Achilles tendons in an overuse rat model. J Orthop Res 2008;26(6):840–6.
- Han JS, Bird GC, Li W, Neugebauer V. Computerized analysis of audible and ultrasonic vocalizations of rats as a standardized measure of pain-related behaviour. Neurosci Meth 2005;141:261–9.

- Khan KM, Cook JL, Bonar F, Harcourt P, Astrom M. Histopathology of common tendinopathies. Update and implications for clinical management. Sports Med 1999;27(6):393–408.
- Khan MH, Li Z, Wang JH. Repeated exposure of tendon to prostaglandin-E2 leads to localized tendon degeneration. Clin J Sport Med 2005;15(1):27–33.
- Lui PP, Fu SC, Chan LS, Hung LK, Chan KM. Chondrocyte phenotype and ectopic ossification in collagenase-induced tendon degeneration. J Histochem Cytochem 2009;57(2):91–100.
- Marxen S, Lacerda Neto JC, Canola JC, Moraes JRE, Ribeiro G. Intralesional polysulphated glycosaminoglycan as treatment of equine collagenase induced tendinitis: clinical, ultrasonographic and histopathologic evaluation. Arq Bras Med Vet Zootec 2004;56(6):701–8.
- Messner K, Wei Y, Andersson B, Gillquist J, Räsänen T. Rat model of Achilles tendon disorder: a pilot study. Cells Tissues Organs 1999;165:30–9.
- Neugebauer V, Han JS, Adwanikar H, Fu Y, Ji GC. Techniques for assessing knee joint pain in arthritis. Mol Pain 2007;3:8.
- Riley GP. The pathogenesis of tendinopathy. A molecular perspective. Rheumatology 2004;43:131–42.
- Rolf CG, Fu SC, Pau A, Wang W, Chan B. Increased cell proliferation and associated expression of PDGFRbeta causing hypercellularity in patellar tendinosis. Rheumatology 2001;40(3):256–61.
- Simjee SU, Pleuvry BJ, Coulthard P. Modulation of the gait deficit in arthritic rats by infusions of muscimol and bicuculline. Pain 2004;109(3):453–60.
- Stone D, Green C, Rao U, Aizawa H, Yamaji T, Niyibizi C, et al. Cytokineinduced tendinitis: a preliminary study in rabbits. J Orthop Res 1999;17(2): 168-77.
- Van Iersel MB, Rikkert M, Borm GF. A method to standardize gait and balance variables for gait velocity. Gait Posture 2007;26:226–30.
- Varejao ASP, Cabrita AM, Meek MF, Bulas-cruz J, Gabriel RC, Filipe VM, et al. Motion of the foot and ankle during the stance phase in rats. Muscle Nerve 2002;26:630–5.
- Vrinten DH, Hamers FFT. CatWalk automated quantitative gait analysis as a novel method to assess mechanical allodynia in the rat; a comparison with von Frey testing. Pain 2003;102:203–9.
- Williams IF, McCullagh KG, Goodship AE, Silver IA. Studies on the pathogenesis of equine tendonitis following collagenase injury. Res Vet Sci 1984;36(3): 326–38.